

0040-4020(94)00600-8

Partial Synthesis of Major Human Metabolites of Docetaxel

Alain Commerçon*, Jean-Dominique Bourzat, Daniel Bézard and Marc Vuilhorgne

Rhône-Poulenc Rorer S.A.- Centre de Recherches de Vitry-Alfortville 13, Quai Jules Guesde - BP14 - 94403 Vitry-sur-Seine (France)

Abstract: The structures and synthesis of the four major human metabolites of docetaxel are reported. These metabolites, *i.e.* compounds 3, 4a,b and 5, are side-chain oxidation derivatives. They were prepared by partial synthesis from the previously described amino-taxoid 8 using mixed-carbonates as acylation reagents. *In vitro* biological activities are detailed.

Taxoids are new anticancer agents that interfere with the microtubule-tubulin system in eukariotic cells¹. Docetaxel 1 (Taxotere[®]), like the related natural analog paclitaxel 2 (Taxol[®]), are currently considered as the most exciting leads of this new series of antitumor drugs. These two taxoids are currently in clinical trials for various types of cancer such as breast, ovarian and non-small cell lung cancers².



R₁ = tBuOCO, R₂ = H (docetaxel)
 R₁ = C₆H₅CO, R₂ = Ac (paclitaxel)

In spite of widening clinical use, little information has been reported thus far on the biological fate of paclitaxel and docetaxel.

Paclitaxel metabolism in animals has been previously studied by Monsarrat *et al.*³⁻⁵. These authors showed that no metabolites of paclitaxel were detected in rat urine, and only 10% of the injected paclitaxel was recovered in urine over a 24-hr period. In contrast, 20% and 40% of the injected drug was recovered in the bile as unchanged paclitaxel and metabolites, respectively. Biotransformation leads to products oxidized at different sites such as the meta-position of the benzoate at C-2 and the para-position of the phenyl group at C-3'. Recently, *in vitro* metabolism of paclitaxel in freshly isolated rat hepatocytes has been investigated⁶. Two metabolites were separated and shown to be monohydroxylated on the side chain and the baccatin core. These results are in agreement with the findings made by Monsarrat *et al.* in the rat bile.

Regarding the metabolism of paclitaxel in humans, it has been reported that small amounts of unchanged drug are found in urine⁷. As in rats, paclitaxel (3%) and its metabolites (17%) were excreted in human bile until 24 hours post-treatment^{5,6}. Five metabolites were detected and three of them were identified as products of mono- and dihydroxylation at the para-position of the phenyl group at C-3' and/or at the C-6 position⁸ of the baccatin moiety. The same monohydroxylated metabolites were also generated *in vitro* by human liver microsomes⁹.

A. COMMERÇON et al.

The principal structural difference between docetaxel 1 and paclitaxel 2 is the presence of a tertbutoxycarbonyl group instead of a benzoyl group on the nitrogen atom of the side-chain. This structural dissimilarity might induce different metabolic transformations. In the course of docetaxel clinical studies, an investigation of human docetaxel metabolites has been conducted. Several metabolites were isolated and structurally characterized in human feces¹⁰. Metabolism of docetaxel in humans leads mainly to side-chainoxidative products. The major isolated metabolites are the side-chain-hydroxylated compound 3, the oxazolidine-type compounds 4 (2 epimers, 4a and 4b) and 5 as well as, in much lower concentrations, the corresponding 7-epimerized compounds of 3-5¹¹.



As metabolites could be very toxic species, thus we synthesized these compounds in order to assess their biological and toxicological effects. This was undertaken using suitable acylation reagents with amino taxoid **8** which is available from 10-deacetyl baccatin III¹².

Preparation of metabolite 3 was achieved as depicted in Scheme 1. Since the taxane skeleton is known to be sensitive to acidic and basic conditions¹³, we chose for the additional hydroxy function a protective group able to be cleaved under either weakly acidic or oxidizing conditions. We selected the para-methoxybenzyl protective group which fulfils both criteria. Thus tertiary alcohol 6 was obtained in 64% yield from the corresponding chlorohydrin and para-methoxybenzyl alcohol under alkylating conditions. Subsequent reaction of 6 with 1,2,2,2-tetrachloroethyl chloroformate afforded the mixed-carbonate 7 (50% yield after filtration over silica gel). By analogy to the work of Barcelo *et al.*¹⁴ with other mixed-carbonates (1,2,2,2-tetrachloroethyl and tertiary alkyl), carbonate 7 was reacted with amino-taxoid 8 in pyridine at room temperature to give within 7h carbamate 9 in 53% yield. Deprotections can be carried out in any order but the best overall yield was obtained by initial removal of the para-methoxybenzyl group with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) in the presence of water (75% yield) and final cleavage of the trichloroethoxycarbonyl (Troc) groups with zinc in acetic acid (60% yield).



Reagents: i) NaH, DMF, r.t., 48h, 64%. ii) pyridine, CH₂Cl₂, r.t., 6h, 50%. iii) pyridine, r.t., 7h, 53%. iv) DDQ, CH₂Cl₂, H₂O, r.t., 1h, 75%. v) Zn, AcOH, MeOH, 60°C, 15min, 60%.

Metabolites 4a,b likely come from metabolite 3 via a subsequent biochemical oxidation of the hydroxylated carbamate moiety. The putative aldehydic intermediate should be able to cyclize to give the two epimeric forms of hydroxyoxazolidinones 4. We attempted the oxidation of 3 as a 2'-O-triethylsilyl-7,10-O-diTroc-protected compound. However in all cases we were unable to isolate any oxidation products of the hydroxy-carbamate moiety. So we undertook a partial synthesis based on an approach, similar to that outlined above, with another suitable mixed-carbonate.

Alcohol 11 was prepared by condensation of methylmagnesium bromide with ethyl diethoxyacetate according to a known procedure¹⁵ (66% yield). Formation of carbonate 12 by reaction with 1,2,2,2-tetrachloroethyl chloroformate in pyridine was achieved in nearly quantitative yield. Acylation of amino-taxoid 8 was performed at room temperature and the reaction was quenched after 24h to give the hemi-acetal 13 (in some experiments the acetal was recovered instead of the hemi-acetal).



Reagents: i) MeMgBr, THF, 10°C, 66%. ii) pyridine, CH₂Cl₂, r.t., 48h, 100% crude. iii) pyridine, CH₂Cl₂, r.t., 24h, 58%. iv) HCOOH, H₂O, AcOEt, r.t., 24h, 61%. v) Zn, AcOH, MeOH, 60°C, 15min., 53% then separation by HPLC.

Hydrolysis of the hemi-acetal was carried out with formic acid at room temperature to give both cyclized epimers in nearly equal amounts (60% total yield)¹⁶. Final cleavage of the Troc groups was performed on the mixture of epimers with zinc in acetic acid. The recovered crude material was purified on silica gel and the two epimeric forms 4a (first epimer eluted) and 4b were separated in 53% yield (ratio 44:56). We have not been able thus far to determine the configuration at the newly formed stereocenter of these two metabolites (efforts of characterization by X-ray crystallographic analysis are in progress).



The last metabolite, *i.e.* compound 5, is likely produced from acid 15^{10} formed by further biochemical oxidation. We were able to obtain by partial synthesis acid 15^{17} . This compound has proved to be rather unstable, cyclizing easily to give the oxazolidinedione derivative 5.

Scheme 2

Scheme 3



Reagents: i) pyridine, CH₂Cl₂, r.t., 24h, 70% crude. ii) aq. K₂CO₃ (5M), THF, r.t., 24h, 78%. iii) Zn, AcOH, MeOH, 60°C, 15min., 53%.

A direct access to compound 5 was realized as follows. The mixed-carbonate 16 was prepared from methyl 2-hydroxyisobutyrate and 1,2,2,2-tetrachloroethyl chloroformate in pyridine (70% yield). Reaction of carbonate 16 with amino-taxoid 8 in a mixture of THF and potassium carbonate (5M) in water led directly to the oxazolidinedione derivative 17 (78% yield)¹⁸. Cleavage of the Troc groups under standard conditions gave metabolite 5 in 53% yield.

The human metabolites have been biologically evaluated in *in vitro* experimental models. Metabolites 4a, 4b and 5 are poor inhibitors of depolymerization in the microtubule disassembly assay¹⁹ (50T, 2.4T and 100T respectively, T being the IC₅₀ value for paclitaxel in the same assay). Metabolite 3 retains a high level of activity in the microtubule disassembly assay (0.65T). This is in good agreement with its close structural analogy with docetaxel. However 3 showed diminished cytotoxicity against P388 leukemia cells²⁰ with an IC₅₀ of 0.45 µg/ml (IC₅₀ for docetaxel was 0.04 µg/ml) while metabolites 4a, 4b and 5 proved to be completely inactive in the same cytotoxicity assay (IC₅₀ >10 µg/ml).

The chemical reactivities observed during the preparation of these major human metabolites of docetaxel are in agreement with the putative metabolic process depicted in Scheme 4. The biochemical sequence likely starts from an hydroxylation of the tert-butoxycarbonyl moiety (metabolite 3) followed by the oxidation of the newly formed hydroxy function to give metabolites 4a,b. We observed by chemical synthesis that the openaldehydic form of compounds 4a,b could not be isolated while both cyclized forms were recovered. Further metabolic oxidation of the putative intermediate aldehydic metabolite leads to the oxazolidine-dione derivative (metabolite 5). Based on our chemical observations, it is postulated that carboxylic acid 15 might be the real biochemical precursor of compound 5 although a biochemical oxidation of hydroxyoxazolidines 4a,b cannot be ruled out. We were also able to detect compound 15 at very low concentrations in human feces using LC-MS experiments¹⁰.



Acknowledgements: We thank Drs. C. Combeau, J.F. Riou, M-C. Bissery, P. Vrignaud and F. Lavelle for biological evaluation, Dr. C. Gaillard for the human metabolites isolation and Dr. C.J. Burns for critical reading of the manuscript.

Experimental Section

NMR spectra were determined with either a Bruker AC-200, AC-300 or AM-400 spectrometer. Chemical shifts (δ) are in ppm relative to TMS (0.00). Mass spectra were produced using FINNIGAN 3300 (70ev; Electron Impact), VG AUTOSPEC (LSIMS Liquid Secondary Ion Mass Spectrometry; matrix: NBA *m*-Nitro Benzyl Alcohol); SCIEX API III (Electrospray); FINNIGAN TSQ45 (DCI; reactant gas: NH₃). Silica gel Merck 60 (0.063-0.20 mm) was used for column chromatography. Preparative thin layer chromatography was carried out on Silica gel Merck 60S-254 plates coated with 0.5 mm layer of silica gel. 2-Methyl-1-(4-methoxyphenyl)methoxy-2-propanol, 6: A solution of 10.9 g (0.1 mol) of 1-chloro-2methyl-2-propanol in 20 mL of DMF was added dropwise at 10°C over 0.5 h to 9.6 g (0.2 mol) of NaH (50% dispersion in oil) in 120 mL of DMF. After 2 h of additional stirring at 20°C, a mixture of 12.5 mL (0.1 mol) of 4-methoxy-benzylalcohol in 20 mL of DMF was added dropwise over 0.5 h. The reaction mixture was stirred at room temperature for 2 days and then heated at 60°C for 8 h to complete the reaction. Then the reaction mixture was poured in 1.2 L of water. The pH was adjusted to 5 by adding 175 mL of hydrochloric acid (1N). The aqueous solution was extracted with three portions of 250 mL of CH₂Cl₂. The solvent was dried with MgSO₄ and evaporated. The crude product was purified on 500 g of silica gel with CH₂Cl₂. This gave 13.6 g (64%) of compound 6 as a pale yellow oil. ¹H-NMR (CDCl₃, 200MHz); δ : 1.24 [s, 6H: -O-C(CH₃)₂], 2.42 (s, 1H: -OH), 3.30 (s, 2H: -OCH₂-), 3.83 (s, 3H: Ar-OCH₃), 4.52 (s, 2H: Ar-CH₂O-), 6.94 (d, J= 8.5 Hz, 2H: Aromatics in ortho of -OCH₃), 7.30 (d, J= 8.5 Hz, 2H: Aromatics in meta of -OCH₃). MS(EI): M/z = 210: M^+ , 138 = M^+ . -OtBu.

2-[2-Methyl-1-(4-methoxyphenyl)methoxy]propyl-1',2',2',2'-tetrachloroethyl-carbonate, 7: By analogy to ref.¹⁴, to a mixture of 5.88 g (28 mmol) of alcohol 6 in 40 mL of CH_2Cl_2 was added at 5°C (ice bath) a solution of 7.59 g (30.8 mmol) of 1,2,2,2-tetrachloroethyl-chloroformate in 40 mL of CH_2Cl_2 . Then 2.43 g (30.8 mmol) of pyridine in 20 mL of CH_2Cl_2 were added dropwise over 0.5 h.. The reaction mixture was stirred for 6 h at 5°C and then washed by three portions (3x50 mL) of water. The organic solvent was dried with MgSO₄ and evaporated. The crude product was filtered on 50 g of silica gel with $CH_2Cl_2/cyclohexane$: 1/1 to give 6.1 g (50%) of 7 as a pale yellow oil used freshly prepared and without further purification.

N-De-t-butoxycarbonyl-N-2-[2-Methyl-1-(4-methoxyphenyl)methoxy]propyloxycarbonyl-7,10-OdiTroc-docetaxel, 9: A solution of 3.36 g (8 mmol) of mixed-carbonate 7 in 25 mL of pyridine was added at 5°C (ice bath) to 4.24 g (4 mmol) of 10-deacetyl-7,10-diTroc-baccatin III 8 in 25 mL of pyridine. The reaction mixture was stirred 8 h at 5°C then concentrated in vacuo. The crude product was chromatographed on 250 g of silica gel with CH₂Cl₂ to give 2.1 g (41 %) of pure compound 9 as a white foam followed by 1.1 g (26%) of recovered 8. ¹H-NMR (CDCl₃, 400MHz); δ: 1.19 (s, 3H: -CH₃ 16 or 17), 1.25 (s, 3H: -CH₃ 16 or 17), 1.36 [s, 6H: -O-C(CH₃)₂], 1.88 (s, 3H: -CH₃ 19), 1.96 (s, 3H: -CH₃ 18), 2.08 [ddd, J= 16, 11 and 2 Hz, 1H: -(CH)-H 6], 2.31 (d, J= 9 Hz, 2H: -CH2- 14), 2.40 (s, 3H: -COCH3), 2.63 [m, 1H: -(CH)-H 6], 2.92 (bb, 1H: -OH in 2'), 3.46 (s, 2H: -OCH2-), 3.80 (s, 3H: Ar-OCH3), 3.91 (d, J= 7 Hz, 1H: -H 3), 4.19 [d, J= 8 Hz, 1H: -(CH)-H 20], 4.34 [d, J= 8 Hz, 1H: -(CH)-H20], 4.41 (ab, J= 12 Hz, 2H: Ar-CH₂O-), 4.61 and 4.93 (2d, J= 12 Hz, 1H each: -OCOO-CH2-CCl3 in 7), 4.65 (bs, 1H: -H 2'), 4.78 (limit AB, 2H: -OCOO-CH2-CCl3 in 10), 4.96 (bd, J= 10 Hz, 1H: -H 5), 5.26 (bd, J= 10 Hz, 1H: -H 3'), 5.52 (d, J= 10 Hz, 1H: -CONH-), 5.55 (dd, J= 11 and 7.5 Hz, 1H: -H 7), 5.70 (d, J= 7 Hz, 1H: -H 2), 6.22 (bt, J= 9 Hz, 1H: -H 13), 6.24 (s, 1H: -H 10), 6.84 (d, J= 8.5 Hz, 2H: Aromatics in ortho of -OCH₃), 7.20 (d, J= 8.5 Hz, 2H: Aromatics in meta of -OCH₃), from 7.30 to 7.45 (m, 5H: -C₆H₅ in 3'), 7.51 [t, J= 8 Hz, 2H: -OCOC₆H₅ (-H 3 and -H 5)], 7.63 [t, J= 8 Hz, 1H: -OCOC6H5(-H4)], 8.11 [d, J= 8 Hz, 2H: -OCOC6H5 (-H 2 and -H 6)]. MS(LSIMS): M/z = 1292: MH+.

N-De-t-butoxycarbonyl-N-2-(1-hydroxy-2-methyl) propyloxycarbonyl-7,10-O-diTroc-docetaxel, 10: To a solution of 5 g (3.86 mmol) of compound 9 in 75 mL of CH₂Cl₂ were added successively 1.5 mL of water and 2.6 g (11.6 mmol) of DDQ. The black reaction mixture was stirred at room temperature for 1 h. The insoluble material was filtered off and the organic solution was washed successively twice with 50 mL of saturated aqueous NaHCO₃ solution and twice with 50 mL of water. The organic solution was dried with MgSO₄ and evaporated. The crude product was purified on 130 g of silica gel with CH₂Cl₂/MeOH : 99/1 to give 3.4 g (75%) of 10 as a white foam. ¹H-NMR (CDCl₃, 400MHz); δ : 1.22 (s, 3H: -CH₃ 16 or 17), 1.30 [s, 9H: -CH₃ 16 or 17 and -O-C(CH₃)₂], 1.88 (s, 3H: -CH₃ 19), 1.99 (s, 3H: -CH₃ 18), 2.09 [ddd, J= 15, 11 and 2 Hz, 1H: -(CH)-H 6], 2.30 and 2.37 (2 dd, J= 16 and 9 Hz, 1H each: -CH₂- 14), 2.43 (s, 3H: -COCH₃), 2.57 (s, 1H: -OH in 1), 2.63 [m, 1H: -(CH)-H 6], 3.29 (d, J= 5 Hz, 1H: -OH in 2'), 3.40 [dd, J= 14 and 5 Hz, 1H: -O-(CH)H]], 3.59 [dd, J= 14 and 7 Hz, 1H: -O-(CH)H]], 3.86 (dd, J= 7 and 5 Hz, 1H: -CH₂-OH), 3.92 (d, J= 7 Hz, 1H: -H 3), 4.22 [d, J= 8.5 Hz, 1H: -(CH)-H 20], 4.71 (dd, J= 5 and 1 Hz, 1H: -(CH)-H 20], 4.61 and 4.92 (2d, J= 12.5 Hz, 1H each: -OCOO-CH₂-CCl₃ in 7), 4.71 (dd, J= 5 and 1 Hz, 1H: -H 2'), 4.79 (limit AB, J= 11 Hz, 2H: -OCOO-CH₂-CCl₃ in 10), 4.97 (bd, J= 9 Hz, 1H: -H5), 5.33 (bd, J= 10 Hz, 1H: -H 3'), 5.57 (dd, J= 11 and 7.5 Hz, 1H: -H 7), 5.63 (d, J= 10 Hz, 1H: -CONH-), 5.74 (d, J= 7 Hz, 1H: -H 2), 6.24 (s, 1H: -H 10), 6.31 (bt, J= 9 Hz, 1H: -H 13), from 7.30 to 7.50 (m, 5H: -C₆H₅ in 3'), 7.51 [t, J= 8 Hz, 2H: -OCOC₆H₅ (-H 3 and -H 5)], 7.62 [t, J= 8 Hz, 1H: -OCOC₆H₅ (-H 4)], 8.12 [d, J= 8 Hz, 2H: -OCOC₆H₅ (-H 2 and -H 6)].

N-De-t-butoxycarbonyl-N-2-(1-hydroxy-2-methyl) propyloxycarbonyl-docetaxel, **3**: To a solution of 3.4 g (2.89 mmol) of 7,10-O-diprotected compound **10** in a mixture of 68 mL of acetic acid and 68 mL of methanol was added at 60°C and under vigorous stirring 6.8 g of zinc powder. Stirring was maintained for 15 min then the reaction mixture was cooled to room temperature, filtered and concentrated in vacuo. The crude product was chromatographed on 90 g of silica gel with $CH_2Cl_2/MeOH : 95/5$ to give 1.2 g (60%) of metabolite **3** as a white foam, $[\alpha]_D^{20}$ -32.8 (c 0.519, MeOH); ¹H-NMR (CDCl₃, at temperature of 323°K, 400MHz); δ : 1.16 (s, 3H: -CH₃ 16 or 17), 1.28 (s, 3H: -CH₃ 16 or 17), 1.31 and 1.33 [s, 6H: -O-C(CH₃)₂], 1.78 (s, 3H: -CH₃ 19), 1.87 [m, 1H: -(CH)-H 6], 1.89 (s, 3H: -CH₃ 18), 2.26 and 2.33 (2 dd, J= 15 and 9 Hz, 1H each: -CH₂- 14), 2.38 (s, 3H: -COCH₃), 2.59 [m, 1H: -(CH)-H 6], 3.26 (bs, 1H: -OH in 2'), 3.44 and 3.60 (2d, J= 14 Hz, 1H each: -CH₂O-), from 3.50 to 3.70 (bb, 1H: -OH), 3.94 (d, J= 7 Hz, 1H: -H 3), 4.19 (bs, 1H: -OH in 10), 4.23 (m, 1H: -H 7), 4.25 [d, J= 8.5 Hz, 1H: -(CH)-H 20], 4.33 [d, J= 8.5 Hz, 1H: -(CH)-H 20], 4.67 (bb, 1H: -H 2'), 4.97 (dd, J= 10 and 2 Hz, 1H: -H 5), 5.21 (s, 1H: -H 10), 5.31 (bd, J= 10 Hz, 1H: -H 3'), 5.62 (d, J= 10 Hz, 1H: -CONH-), 5.72 (d, J= 7 Hz, 1H: -H 2), 6.29 (bt, J= 9 Hz, 1H: -H 13), from 7.30 to 7.45 (m, 5H: -C₆H₅ in 3'), 7.50 [t, J= 8 Hz, 2H: -OCOC₆H₅ (-H 3 and -H 5)], 7.61 [t, J= 8 Hz, 1H: -OCOC₆H₅ (-H 4)], 8.12 [d, J= 8 Hz, 2H: -OCOC₆H₅ (-H 2 and -H 6)]. MS(LSIMS): M/z = 824: MH⁺.

2-(1,1-Diethoxy-2-methyl-propyl)-1',2',2',2'-tetrachloroethyl-carbonate, 12: By analogy to ref.¹⁴, with the use of a similar procedure as for 7, 0.81 g (50 mmol) of 2-(1,1-diethoxy-2-methyl)propanol 11 gave 1.8 g (100%) of carbonate 12 as a pale yellow oil used freshly prepared in the next step.

N-De-t-butoxycarbonyl-N-2-(1-ethoxy-1-hydroxy-2-methyl)propyloxycarbonyl-7,10-O-diTroc-docetaxel, 13: With the use of a similar procedure as for **8**, 1.06 g (1 mmol) of amino-taxoid **6** and 0.41 g (1.1 mmol) of carbonate **12** in 10 mL of pyridine led after purification on 50 g of silica gel with $CH_2Cl_2/MeOH$: 98/2 0.53 g of compound **13** as a white foam. ¹H-NMR (CDCl₃, at temperature of 323°K, 400MHz); δ : from 1.10 to 1.40 (m: -CH₃ of the ethyl group), 1.21 (s, 3H: -CH₃ 16 or 17), 1.29 (s, 3H: -CH₃ 16 or 17), 1.35 [s, 6H: -O-C(CH₃)₂], 1.70 (s, 1H: -OH in 1), 1.88 (s, 3H: -CH₃ 19), 1.98 (s, 3H: -CH₃ 18), 2.08 [ddd, J= 16, 11 and 2 Hz, 1H: -(CH)-H 6], 2.33 (limit ab, 2H: -CH₂- 14), 2.38 (s, 3H: -COCH₃), 2.64 [m, 1H: -(CH)-H 6], 2.81 (d, J= 11 Hz, 1H: -OH), 3.32 (d, J= 5.5 Hz, 1H: -OH in 2), from 3.50 to 3.90 (m: -OCH₂- of the ethyl group), 3.94 (d, J= 7 Hz, 1H: -H 3), 4.21 [d, J= 8 Hz, 1H: -(CH)-H 20], 4.33 [d, J= 8 Hz, 1H: -(CH)-H 20], 4.62 and 4.91 (2d, J= 13 Hz, 1H each: -OCOO-CH₂-CCl₃ in 7), 4.64 (dd, J= 5 and 2.5 Hz, 1H: -H 2'), 4.79 (limit AB, J= 14 Hz, 2H: -OCOO-CH₂-CCl₃ in 10), 4.87 (d, J= 11 Hz, 1H: -O-CH-O-), 4.97 (bd, J= 9.5 Hz, 1H: -H 5), 5.25 (bd, J= 10 Hz, 1H: -H 3'), 5.45 (d, J= 10 Hz, 1H: -CONH-), 5.55 (dd, J= 11 and 7 Hz, 1H: -H 7), 5.72 (d, J= 7 Hz, 1H: -H 2), 6.25 (bt, J= 9 Hz, 1H: -H 13), 6.28 (s, 1H: -H 10), from 7.30 to 7.45 (m, 5H: -C₆H₅ in 3'), 7.51 [t, J= 8 Hz, 2H: -OCOC₆H₅ (-H 3 and -H 5)], 7.62 [t, J= 8 Hz, 1H: -OCOC₆H₅ (-H 4)], 8.12 [d, J= 8 Hz, 2H: -OCOC₆H₅ (-H 2 and -H 6)]. MS(LSIMS) : M/z = 1198: MH⁺- H₂O.

3'-De-t-butoxycarbonylamino-3'-[3-(5,5-dimethyl-4-hydroxy-2-oxo-1,3-oxazolidinyl)]-7,10-OdiTroc-docetaxel, 14, as a mixture of diastereomers: To a mixture of 0.22 g of compound 13 in 5 mL of ethylacetate was added a solution of 2 mL of formic acid in 2 mL of water. The reaction mixture was stirred at room temperature for 24 h then diluted with water and extracted with ethyl acetate. The organic solution was dried with MgSO₄ and evaporated. The crude product was purified on 10 g of silica gel with CH₂Cl₂/MeOH : 97/3 to give 0.183 g (87%) of 14 (mixture 44:56 of diastereomers) as a white foam. ¹H-NMR (CDCl₃, at temperature of 323°K, 400MHz); δ : 1.18, 1.21, 1.24, 1.27, 1.33, 1.40 and 1.42 [7s, 12H: -CH₃ 16, -CH₃ 17 and -O-C(CH₃)₂], from 1.80 to 2.20 [m, 1H: -(CH)-H 6], 1.84 and 1.86 (2s, 3H: -CH₃ 19), 2.01 and 2.05 (2s, 3H: -CH₃ 18), 2.10 and 2.19 (2 dd, J= 14 and 9 Hz, 1H: -CH₂- 14 of one diastereoisomer], 2.24 and 2.30 (2s, 3H: -COCH₃), 2.38 (d, J= 9 Hz, 1H: -CH₂- 14 of the other diastereoisomer), 2.62 [m, 1H: -(CH)-H 6], 3.90 (d, J= 7 Hz, 1H: -H 3), 4.16 and 4.30 (2d, J= 8 Hz, 1H: -CH₂- 20 of one diastereoisomer), 4.22 and 4.31 (2d, J= 8.5 Hz, 1H: -CH₂- 20 of the other diastereoisomer), 4.40 and 4.78 (2s, 1H: CH-O), 4.60 and 4.90 (2d, J= 12 Hz, 1H each: -OCOO-CH₂-CCl₃ in 7), 4.72 (bs, 0.5 H: -H 2' of one diastereoisomer), 4.79 (limit AB, J= 14 Hz, 2H: $-OCOO-CH_2-CCl_3$ in 10), 4.95 (bd, J= 9.5 Hz, 1H: -H 5), 4.97 (d, J= 6 Hz, 0.5 H: -H 2' of the other diastereoisomer), 5.02 (d, J= 6 Hz, 0.5 H: -H 3' of one diastereoisomer), from 5.50 to 5.60 (m, 1H: -H 7), 5.55 (bs, 1H: -H 3' of the other diastereoisomer), 5.67 and 5.71 (2d, J= 7 Hz, 1H: -H 2), 6.13 and 6.25 (2 bt, J= 9 Hz, 1H: -H 13), 6.26 (s, 1H: -H 10), from 7.30 to 7.70 [m, 8H: $-C_6H_5$ in 3' and $-OCOC_6H_5$ (-H 3, -H 4 and -H 5)], 8.06 and 8.10 [2d, J= 8 Hz, 2H: $-OCOC_6H_5$ (-H 2 and -H 6)].

3'-De-t-butoxycarbonylamino-3'-[3-(5,5-dimethyl-4-hydroxy-2-oxo-1,3-oxazolidinyl)]-docetaxel, 4a and 4b: By analogy to ref.¹⁴, with the use of a similar procedure as for 3, 0.95 g (0.085 mmol) of 7,10-O-diTroc derivative 14 in 2 mL and acetic acid and 2 mL of methanol treated with 0.2 g of zinc powder led, after purification on 10 g of silica gel with $CH_2Cl_2/MeOH : 95/5$, 0.035 g (56%) of compound 4 as a mixture of diastereomers (4a:4b = 44:56). After further purification using preparative TLC ($CH_2Cl_2/MeOH : 9/1$), 4a (first product eluted) and then 4b were obtained as white foams:

4a: $[\alpha]_{D}^{20}$ -13.1 (c 0.610, MeOH); ¹H-NMR (CDCl₃, 400 MHz); δ : 1.13 (s, 3H: -CH₃ 16 or 17), 1.18, 1.26 and 1.42 [3s, 9H: -CH₃ 16 or 17 and -O-C(CH₃)₂], 1.75 (s, 3H: -CH₃ 19), 1.85 [ddd, J=16, 11 and 2 Hz, 1H: -(CH)-H 6], 1.89 (s, 1H: -OH in 1), 1.92 (s, 3H: -CH₃ 18), 2.17 (s, 3H: -COCH₃), 2.31 and 2.37 (dd, J= 16 and 9 Hz, 1H each: -CH₂- 14), 2.53 [m, 1H: -(CH)-H 6], 3.86 (d, J= 7 Hz, 1H: -H 3), 4.20 (d, J= 8.5 Hz, 1H: -(CH)-H 20), 4.23 (dd, J= 11 and 7 Hz, 1H: -H 7), 4.30 [d, J= 8.5 Hz, 1H: -(CH)-H 20], 4.34 (s, 1H: -OH in 10), 4.38 (bs, 1H: CH-O-), 4.97 (dd, J= 10 and 2 Hz, 1H: -H 5), 5.00 (d, J= 2.5 Hz, 1H: -H 2), 5.31 (s, 1H: -H 10), 5.56 (d, J= 2.5 Hz, 1H: -H 3), 5.68 (d, J= 7 Hz, 1H: -H 2), 6.26 (bt, J= 9 Hz, 1H: -H 2), 5.31 (s, 1H: -H 10), 5.56 (d, J= 2.5 Hz, 1H: -H 3), 7.40 [t, J= 8 Hz, 2H: -C₆H₅ in 3' (-H 3 and -H 5)], 7.53 [t, J= 8 Hz, 2H: -OCOC₆H₅ (-H 3 and -H 5)], 7.58 [d, J= 8 Hz, 2H: -OCOC₆H₅ (-H 4)], 8.11 [d, J= 8 Hz, 2H: -OCOC₆H₅ (-H 2 and -H6)]. MS(DCI): M/z = 839: MNH4⁺.

4b: $[\alpha]_D^{20}$ -46.2 (c 0.515, MeOH); ¹H-NMR (CDCl₃, 400MHz); δ : 1.12 (s, 3H: -CH₃ 16 or 17), 1.22 (s, 3H: -CH₃ 16 or 17), 1.34 and 1.44 [3s, 6H: -O-C(CH₃)₂], 1.72 (s, 1H: -OH in 1), 1.75 (s, 3H: -CH₃ 19), 1.85 [ddd, J= 16, 11 and 2 Hz, 1H: -(CH)-H 6], 1.98 (s, 3H: -CH₃ 18), 2.01 and 2.11 (2 dd, J= 16 and 9 Hz, 2H: -CH₂-14), 2.30 (s, 3H: -COCH₃), 2.30 (s, 3H: -COCH₃), 2.57 [m, 1H: -(CH)-H 6], 3.66 (bb, 1H: -OH in 2'), 3.90 (d, J= 7 Hz, 1H: -H 3), 4.15 [d, J= 8 Hz, 1H: -(CH)-H 20], 4.26 (dd, J= 11 and 7 Hz, 1H: -H 7), 4.30 [d, J= 8 Hz, 1H: -(CH)-H 20], 4.31 (bs, 1H: -OH in 10), 4.68 (bs, 1H: CH-O-), 4.93 (dd, J= 10 and 2 Hz, 1H: -H 5), 4.96 (bd, J= 6.5 Hz, 1H: -H 2'), 5.01 (d, J= 6.5 Hz, 1H: -H 3'), 5.24 (s, 1H: -H 10), 5.31 (bb, 1H: -OH), 5.64 (d, J= 7 Hz, 1H: -H 2), 6.14 (bt, J= 9 Hz, 1H: -H 13), 7.33 [t, J= 8 Hz, 1H: -C₆H₅ in 3' (-H 4)], 7.41 [t, J= 8 Hz, 2H: -C₆H₅ in 3' (-H 4)], 7.41 [t, J= 8 Hz, 2H: -OCOC₆H₅ (-H 3 and -H 5)], 7.64 [t, J= 8 Hz, 1H: -OCOC₆H₅ (-H 4)], 8.06 [d, J= 8 Hz, 2H: -OCOC₆H₅ (-H 2 and -H 6)].MS(DCI): M/z = 839: MNH₄⁺.

2-(2-methoxycarbonyl)propyl)-1',2',2',2'-tetrachloroethyl-carbonate, 16: With the use of a similar procedure as for 7, 2.36 g (0.2 mol) of methyl 2-hydroxyisobutyrate gave 4.57 g (70%) of carbonate 16 as white powder (m.p. $<50^{\circ}$ C) used freshly prepared in the next step.

3'-De-t-butoxycarbonylamino-3'-[3-(5,5-dimethyl-2,4-dioxo-1,3-oxazolidinyl)]-7,10-O-diTroc-

docetaxel, 17: To a mixture of 0.264 g (0.25 mmol) of amino-taxoid 8 in 5 mL of THF were added successively 0.082 g (1.1 mmol) of carbonate 16 and 1 mL of an aqueous solution of K_2CO_3 (5M). The reaction mixture was stirred at room temperature for 24 h. Then 15 mL of water were added and extraction was performed with ethyl acetate (3x10 mL). The organic solution was dried with MgSO₄ and evaporated in vacuo. The crude product was purified on 15 g of silica gel with CH₂Cl₂/MeOH : 99/1 to give 0.228 g (78%) of 17 as a white foam. ¹H-NMR (CDCl₃, 400 MHz); δ : 1.17 (s, 3H: -CH₃ 16 or 17), 1.21 (s, 3H: -CH₃ 16 or 17), 1.60 and 1.63 [2s, 3H each: -O-C(CH₃)₂], 1.83 and 2.03 (2 dd, J= 14 and 9 Hz, 1H each: -CH₂-14), 1.84 (s, 3H: -CH₃ 19), 1.98 (s, 3H: -CH₃ 18), 2.05 [ddd, J= 16, 11 and 2 Hz, 1H: -(CH)-H 6], 2.39 (s, 3H: -COCH₃), 2.63 [m, 1H: -(CH)-H 6], 3.85 (d, J= 7 Hz, 1H: -H 3), 4.00 (bb, 1 H: -OH in 2'), 4.14 [d, J= 8 Hz, 1H: -(CH)-H 20], 4.31 [d, J= 8 Hz, 1H: -(CH)-H 20], 4.60 and 4.92 (2d, J= 11 Hz, 1H each: -OCOO-CH₂-CCl₃ in 7), 4.77 (limit AB, J= 12 Hz, 2H: -OCOO-CH₂-CCl₃ in 10), 4.96 (bd, J= 9.5 Hz, 1H: -H 5), 5.26 (bd, J= 8 Hz, 1H: -H 2'), 5.40 (d, J= 8 Hz, 1H: -H 10), 7.31 [t, J= 8 Hz, 1H: -H 7), 5.64 (d, J= 7 Hz, 1H: -H 2), 6.07 (bt, J= 9 Hz, 1H: -H 13), 6.22 (s, 1H: -H 10), 7.31 [t, J= 8 Hz, 1H: -C₆H₅ in 3' (-H 4)], 7.40 [t, J= 8 Hz, 2H: -OCOC₆H₅ (-H 4) and -H 5)], 7.67 [t, J= 8 Hz, 1H: -OCOC₆H₅ (-H 4)], 8.06 [d, J= 8 Hz, 2H: -OCOC₆H₅ (-H 2 and -H 6)]. MS(DCI) : M/z = 1185: MNH4⁺.

3'-De-t-butoxycarbonylamino-3'-[3-(5,5-dimethyl-2,4-dioxo-1,3-oxazolidinyl)]-docetaxel, 5: With the use of a similar procedure as for **3**, 0.15 g (0.128 mmol) of 7,10-O-diTroc derivative **17** in **3** mL of acetic acid and 3 mL of methanol treated with 0.3 g of zinc powder at 60°C led, after purification on 5 g of silica gel with CH₂Cl₂/MeOH : 98/2, 0.055 g (53%) of pure compound **5** as a white foam, $[\alpha]_{D}^{20}$ -49.3 (c 0.531, MeOH); ¹H-NMR (CDCl₃, 400 MHz); $\delta : 1.10$ (s, 3H: -CH₃ 16 or 17), 1.18 (s, 3H: -CH₃ 16 or 17), 1.60 and 1.63 [2s, 3H each: -O-C(CH₃)₂], 1.74 (s, 3H: -CH₃ 19), 1.80 (m, 2H: H 14 and H 6), 1.9 (s, 3H: -CH₃ 18), 1.98 (dd, J= 14 and 9 Hz, 1H: H 14), 2.15 (s, 3H: -COCH₃), 2.60 [m, 1H: H 6], 3.86 (d, J= 7 Hz, 1H: -H 3), 4.02 (bd, 1 H: -OH in 2'), 4.15 to 4.30 [m, 4H: 2xH 20, H 7 and OH 7), 4.94 (bd, J= 9.5 Hz, 1H: -H 5), 5.20 (s, 1H: -H 13), 7.28 [t, J= 8 Hz, 1H: -H 2'), 5.40 (d, J= 8 Hz, 1H: -H 3'), 5.61 (d, J= 7 Hz, 1H: -H 2), 6.04 (bt, J= 9 Hz, 1H: -H 13), 7.28 [t, J= 8 Hz, 1H: -C₆H₅ in 3' (-H 4)], 7.40 [t, J= 8 Hz, 2H: -C₆H₅ in 3' (-H 3 and -H 5)], 7.50 [d, J= 8 Hz, 2H: -C₆H₅ in 3' (-H 2 and -H 6)], 7.53 [t, J= 8 Hz, 2H: -OCCC₆H₅ (-H 3 and -H 5)], 7.65 [t, J= 8 Hz, 1H: -OCOC₆H₅ (-H 4)], 8.06 [d, J= 8 Hz, 2H: -OCCOC₆H₅ (-H 2 and -H 6)]. MS(LSIMS) : M/z = 820: MH⁺.

References and notes:

- 1. a) Schiff P.B., Fant J., Horwitz S.B., Nature, 1979, 665. b) Manfredi J.J., Horwitz S.B., Pharmac. Ther., 1984, 83.
- 2. a) For a recent review on preclinical studies on docetaxel, see: Lavelle F., Guéritte-Voegelein F., Guénard D., *Bull. Cancer*, 1993, 80, 326. b) For clinical results on docetaxel and paclitaxel, see: Rothenberg M.L., *Curr. Opin. Invest. Drugs*, 1993, 2 (12), 1269. c) Paclitaxel has been recently approved by the FDA for ovarian cancer treatment.
- 3. Monsarrat B., Mariel E., Cros S., Garès M., Guénard D., Guéritte-Voegelein F., Wright M., Drug Metab. Dispos., 1990, 18, 895.
- Monsarrat B., Alvinerie P., Garès M., Wright M., Dubois J., Guéritte-Voegelin F., Guénard D., Donehower R., Rowinsky E., Cell. Pharmacol., 1993, 1 (suppl. I), S77.
- 5. Monsarrat B., Alvinerie P., Wright M., Dubois J., Guéritte-Voegelin F., Guénard D., Donehower R., Rowinsky E., J. Natl. Cancer Inst. Monograph., 1994, in press.
- 6. Walle T., Kumar G.N., McMillan J.M., Thornburg K.R., Walle U.K., Biochem. Pharmacol., 1993, 46 (9), 1661.
- 7. Rowinsky E.K., Cazenave I.A., Donehower R.C., J. Natl. Cancer Inst., 1990, 82, 1247.
- 8. Harris J.W., Katki A., Anderson L.W., Chmurny G.N., Paukstelis J.V., Collins J.M., J. Med. Chem., 1994, 37, 706.
- 9. Cresteil T., Monsarrat B., Alvinerie P., Tréluyer J.M., Vieira I., Wright M., Cancer Res., 1994, 54, 386.
- a) Vuilhorgne M. et al., presented at the 207 th ACS National Meeting, San Diego, March 13-17, 1994.
 b) Vuilhorgne M. et al., ACS Symposium Series, in preparation. c) Monegier B., Gaillard C., Sablé S., Vuilhorgne M., Tetrahedron Lett., 1994, in press.
- 11. The epimerization of paclitaxel at C-7 in cell culture medium has been previously observed, see: Ringel I., Horwitz S.B., J. Pharmacol. Exp. Ther., 1987, 242, 692.
- 12. Commerçon A., Bézard D., Bernard F., Bourzat J.D., Tetrahedron Lett., 1992, 33, 5185.
- For very recent reviews about taxoid chemistry, see: Kingston D.G.I., Molinero A.A., Rimoldi J.M., Progress in the Chemistry of Organic Natural Products, 1993, 61, 1; Nicolaou K.C., Dai W.M., Guy R.K., Angew. Chem. Int. Ed. Engl., 1994, 33, 15.
- 14. Barcelo G., Senet J.P., Sennyey G., Bensoam J., Loffet A., Synthesis, 1986, 627; Olofson R.A. Pure Appl. Chem., 1988, 60, 1715.
- 15. Avy M.A., Bull. Soc. Chim. Fr., 1931, 49, 12.
- 16. For another example of preparation of 5-hydroxy-2-oxo-1,3-oxazolidines, see: Kano S., Yuasa Y., Yokomatsu T., Shibuya S., Chem. Lett., 1983, 1475.
- 17. Bourzat J.D. et al., unpublished results.
- For a previous preparation of oxazolidine-2,4-diones, see: Rekker R.F., Verleur H., Nauta W.Th., Rec. Trav. Chim. Pays-Bas, 1951, 70, 5; Rekker R.F., Faber A.C., Tom D.H.E., Verleur H., Nauta W.Th., Rec. Trav. Chim. Pays-Bas, 1951, 70, 113.
- 19. Chauvière G., Guénard D., Picot F., Sénilh V., Potier P., C. R. Acad. Sciences, Série II, 1981, 293, 501.
- 20. Riou J.F., Naudin A., Lavelle F., Biochem. Biophys. Res. Commun., 1992, 187, 164.

(Received in Belgium 7 March 1994; accepted 19 May 1994)